

Influence of Fluctuating Water Levels on Mercury Concentrations in Adult Walleye

T. M. Selch · C. W. Hoagstrom · E. J. Weimer ·
 J. P. Duehr · S. R. Chipps

Received: 20 December 2006 / Accepted: 30 May 2007 / Published online: 7 July 2007
 © Springer Science+Business Media, LLC 2007

Keywords Mercury · Monitoring · Walleye ·
 Water level fluctuations

Atmospheric deposition of mercury (Hg) has been shown to be a significant source of Hg on the landscape (Rada et al. 1989; Swain et al. 1992) and is believed to contribute to increased Hg concentrations in aquatic food webs (Sorensen et al. 1994; Edwards et al. 1999). Methylmercury (MeHg) is the organic, bioavailable form of Hg that accumulates to toxic levels in top-level predators in aquatic systems (Suedel et al. 1994). Although limnological conditions in lakes and rivers can affect Hg methylation and concentrations in fishes, these relationships often vary among water bodies and fish species (McMurtry et al. 1989; Bodaly et al. 1993). Thus, regional studies are needed

to identify mechanisms of local MeHg production and factors associated with Hg contamination in fishes.

The Prairie Pothole Region of North America has a unique variety of natural wetlands and glacial lakes that are important for fish, shorebirds, waterfowl, and humans. Cyclical climate, characterized by extended wet–dry periods, cause high variation in water surface area within the region (Rosenberry 2003). For example, consecutive years of high precipitation during the mid-1990s caused dramatic surface area increases in many glacial lakes and wetlands of eastern South Dakota. After lake levels increased, several fish populations were found to contain elevated Hg concentrations ($>1 \mu\text{g/g}$), prompting local officials to post fish consumption advisories (South Dakota Game, Fish and Parks, 2006).

The discovery of elevated Hg concentrations in fishes was surprising because (1) there were no apparent point-source inputs of Hg, and (2) the limnological conditions of most of these lakes (eutrophic, high pH) generally do not favor Hg methylation or bioaccumulation (Grieb et al. 1990; Pickhardt et al. 2002). Moreover, lakes that experienced large increases in surface area generally contained fast growing fish populations, a situation that usually lowers Hg concentration due to growth dilution (Rodgers and Qadri 1982; MacCrimmon et al. 1983). In this study, we document changes in lake surface area for glacial lakes in the Prairie Pothole Region and relate this to Hg concentration in adult walleye (*Sander vitreus*). Although a similar phenomenon is known to occur in reservoirs with fluctuating water levels (Jackson 1988; Snodgrass et al. 2000; Sorensen et al. 2005), widespread effects of surface area changes on Hg concentrations in fishes have not been documented in natural, glacial lakes. We hypothesized that increases in lake surface area enhanced

T. M. Selch (✉) · S. R. Chipps
 U.S. Geological Survey South Dakota Cooperative Fish and
 Wildlife Research Unit, Department of Wildlife and Fisheries
 Sciences, South Dakota State University, Box 2140B,
 Brookings, SD 57007-1696, USA
 e-mail: Trevor.Selch@sdstate.edu

C. W. Hoagstrom
 Department of Zoology, Weber State University,
 2505 University Circle, Ogden, UT 84408-2505, USA

E. J. Weimer
 Ohio Department of Natural Resources, Division of Wildlife,
 Sandusky Fisheries Unit, 305 E. Shoreline Drive, Sandusky,
 OH 44870, USA

J. P. Duehr
 Westwood Professional Services, 7699 Anagram Drive,
 Eden Prairie, MN 55344-7310, USA

Hg methylation and resulted in elevated fish Hg concentrations.

Materials and Methods

We studied 18 lakes within the Northern Glaciated Plains Ecoregion of eastern South Dakota (Fig. 1). Lakes in this region range from eutrophic to hypereutrophic and generally do not thermally stratify during summer months (i.e., polymictic mixing cycles). Changes in lake surface area were determined using Landsat 5 imagery (<http://www.sdview.sdstate.edu>) collected during the late 1980s (dry period) and early 2000s (wet period). Lake surface areas, determined from images obtained in 1987 and 2000, were digitized in ArcMap 9.1 to quantify the surface area (ha) of each lake for both time periods. We used regression analysis to assess the relationship between Hg concentrations in walleyes and percent change in surface area (SA) of lakes between wet (2000) and dry (1987) years.

Adult walleyes (350–500 mm total length TL) were collected during summer months (June through August) from 1996 to 2005 using a combination of electrofishing, trap-nets, and experimental gill-nets. Muscle samples (~2 g) obtained from 1996 to 2004 were collected from whole walleye filets. A composite sample was obtained from five similar sized walleyes (50 mm size categories) then homogenized and analyzed for total Hg. Three to six composite samples were obtained from each lake and averaged to quantify walleye Hg concentration. In 2005, eight additional lakes were sampled and walleye filets

(10–15 fish/lake) were analyzed individually for total Hg concentration, and then averaged to determine mean Hg concentration for the lake. All tissue samples were analyzed for total Hg using cold vapor atomic fluorescence spectrometry (Jones et al. 1997; Collin-Hansen et al. 2005; Yu 2005). The standard reference material (SRM) used was National Institute of Standards and Technology (NIST) #2976 muscle tissue. Our SRM contained 54.6 (2.1) ng/g total Hg. Fish samples were spiked at a level of either 0.5, 1.0, or 3.0 ng/g, with a detection limit of 0.02 ng/g. Percent spike recovery in our samples averaged 100.2 (4.6). Minimum detection limits for our fish tissue samples were <0.02 µg/g total Hg.

Information on watershed characteristics and water quality attributes were available for 10 of the 18 lakes we sampled (Stukel 2003). We used these data to explore relationships between walleye Hg concentrations and environmental factors. Variables were tested for homogeneity of variance and normality. Pearson correlations were used to identify significant relationships between individual parameters and walleye Hg levels ($\alpha = 0.05$)

Results and Discussion

Mean Hg concentrations in walleyes varied considerably among lakes, ranging from 0.05 (Pelican lake) to 0.99 µg/g (Bitter and Twin lakes; Table 1). Changes in lake surface area, as determined by difference (i.e., wet year–dry year), ranged from –54 ha (Lake Madison) to +3,683 ha (Waubay Lake). On a percentage basis, Lake Madison had the largest decrease in surface area (4.8%), while Lynn Lake expanded in size by over 300% (Table 1). Percent data were log transformed to correct for normality because we had a right-skewed distribution. The increase in lake SA associated with wet periods of the mid-1990s was significantly related to walleye Hg concentrations ($n = 18$, model $F_{1,17} = 26.0$, $r^2 = 0.62$, $p < 0.0001$; Fig. 2). Analysis of watershed and water quality variables from the ten lakes data set showed that most variables were poorly correlated to walleye Hg concentration, except SA change (Table 2).

Rapid increases in lake levels during the 1990s may be analogous to the “reservoir effect” and explain variation in Hg concentrations among the lakes we studied. Methylation of Hg in newly flooded soils may remain high for 10–15 years post-inundation (Porvari 1998; Bowles et al. 2003), so it is possible that walleye Hg concentrations will remain high for several more years. It appears that atmospheric Hg deposition in eastern South Dakota (Gossman 2003; EPA 2005) is not a trivial contribution and can accumulate in adjacent terrestrial soils and contribute to Hg contamination in aquatic food webs when flooding occurs. In our study, the magnitude of lake surface area expansions

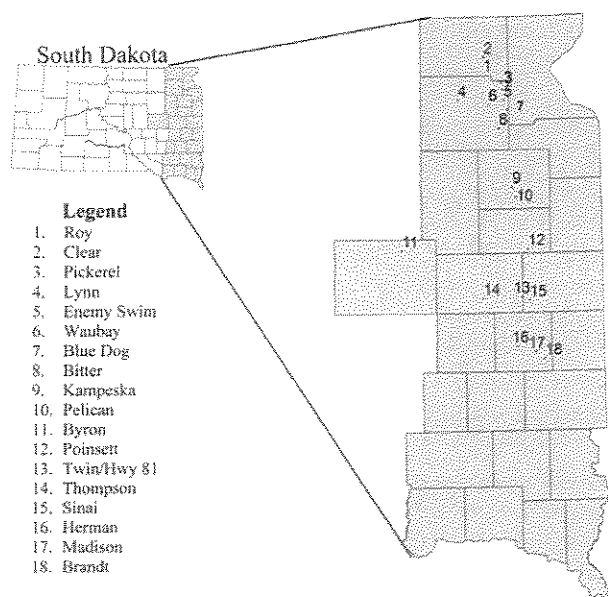


Fig. 1 Location of 18 study lakes in eastern South Dakota

Table 1 Surface area of lakes in 1987 (dry period) and 2000 (wet period), and mean Hg levels in adult walleyes collected from 1996–2005

Lake	Surface area 1987 (ha)	Surface area 2000 (ha)	Percent change in surface area	Walleye Hg ($\mu\text{g/g}$)	Sample collection year
Bitter	1309.9	4405.4	236.3	0.99	2005
Blue Dog	669.9	761.8	13.7	0.21	2000
Brandt	426.6	506.7	18.8	0.18	1998
Byron	751.8	746.9	−0.6	0.19	2005
Clear	468.7	484.7	3.4	0.14	1999
Enemy Swim	884.0	884.0	0.0	0.19	2005
Herman	521.3	502.2	−3.7	0.10	1996
Kampeska	1990.4	2046.8	2.8	0.30	2005
Lynn	157.2	643.4	309.4	0.57	2005
Madison	1109.3	1055.8	−4.8	0.21	1999
Pelican	1124.8	1124.8	0.0	0.05	2005
Pickerel	407.6	407.6	0.0	0.17	2000
Poinsett	3160.9	3160.9	0.0	0.13	1997
Roy	631.4	845.3	33.9	0.11	2005
Sinai	284.2	751.9	164.6	0.43	1996
Thompson	4989.9	5393.4	8.1	0.42	1996
Twin	364.9	1025.9	181.2	0.99	2005
Waubay	3648.2	7331.2	100.9	0.40	2001

The surface area of each lake was determined using Landsat 5 images collected in 1987 and 2000

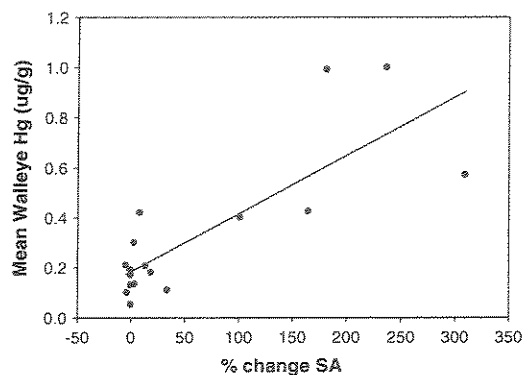


Fig. 2 Relationship between mean walleye Hg concentrations ($\mu\text{g/g}$) for eastern South Dakota glacial lakes and percent change in surface area (SA) between wet (2000) and dry (1987) years [$n = 18$, $F_{(1,17)} = 26.0$, $p < 0.0001$, $r^2 = 0.62$, $y = 0.5296 + 0.372(\log \text{ percent change in surface area})$]

was positively related to Hg concentrations in walleye (Table 1). However, it not known how the duration and/or frequency of wet–dry cycles affect Hg levels in specific water-bodies, because limnological conditions and water cycles vary between lakes.

Increased Hg concentrations measured in fishes following water level fluctuations could be associated with the frequency of inundation (Sorensen et al. 2005). Increased sulfate levels, caused by the drying and rewetting of soils, enhance sulfate reducing bacteria that produce MeHg.

Lakes that endure recurrent annual wet–dry cycles likely experience lower sulfate mobilization than a lake that has not been inundated for many years (Gilmour et al. 2004; Sorensen et al. 2005). Thus, extended wet–dry periods in eastern South Dakota may have resulted in elevated sulfate concentrations that enhance Hg production and availability in these systems (St. Louis et al. 2004).

Productivity of glacial lakes may contribute to Hg concentrations in fish. For example, high walleye growth rates (based on age-3 TL) are typical in many lakes with elevated Hg concentrations (South Dakota Game, Fish and Parks 2006). Fast growing fish populations should result in fish with lower Hg concentrations owing to growth dilution (Norstrom et al. 1976; Olsson 1976; Verta 1990) and high algal productivity (Pickhardt et al. 2002; Essington and Houser 2003); however, based on growth data for age-3 walleyes reported in Stukel (2003), we found that walleye growth was positively correlated with mean Hg concentration ($n = 10$, $r = 0.695$, $p = 0.026$). The surface area changes that appear to enhance Hg methylation also increase the productivity of these systems. As a result, the buffering effect of high fish growth rates and primary productivity (decreased Hg burden per algal cell) may not be realized in natural lakes that increase in surface area because of the link between productivity and Hg accumulation.

Although variables such as pH, alkalinity, surface area, and watershed area explained fish Hg concentrations in

Table 2 Results from Pearson correlation analysis comparing mean walleye Hg concentrations with environmental variables from ten glacial lakes in eastern South Dakota

Parameter	<i>r</i>	<i>p</i>
WsL	-0.450	0.192
Vol:area ratio	0.397	0.257
Chl- <i>a</i> (mg/L)	0.391	0.263
pH	-0.006	0.987
Alkalinity (mg/L)	0.450	0.192
Conductivity (mS/cm)	0.252	0.483
% SA	0.759*	0.011*

pH, alkalinity, conductivity, and percent change in surface area between wet (2000) and dry (1987) years (% SA). Environmental data are from Stukel (2003)

WsL watershed area to lake surface area ratio, Vol:area ratio lake volume to surface area ratio, Chl-*a* chlorophyll-*a* concentration

*Significant correlations ($p < 0.05$)

other studies (McMurtry et al. 1989; Bodaly et al. 1993; Rudd 1995), they did not correlate with walleye Hg levels in our study (Table 2). This may be due to the high methylation rates resulting from organic matter decomposition that followed water level increases in the 1990s. Further, low pH (<7.0) increases microbial methylation of mercury (Wren and MacCrimmon 1983; Grieb et al. 1990; Hakanson 2003), but water pH was relatively high in our study lakes (mean = 8.9 ± 0.04 SE), and may explain why pH did not influence Hg contamination.

Our study lakes were located in close proximity to each other in eastern South Dakota (Fig. 1). Although the region experienced relatively uniform precipitation during the extended wet period of the mid-1990s, some lakes expanded faster and several years earlier than others. Lakes from early water-level expansions may be receding in MeHg production, and reduced MeHg production within a lake should result in lower total Hg concentrations in the resident fish communities. Moreover, many lakes experienced little to no change in surface area between wet and dry years (Table 1). Several of these lakes (i.e., Kampeska, Pelican, and Poinsett) have water control structures that maintain stable water levels in the lake, and generally contain fish with low Hg concentrations.

Recent fish consumption advisories in South Dakota (South Dakota Game, Fish and Parks 2006) indicate that Hg contamination is a concern. Walleye are a popular sport fish in the Prairie Pothole Region and their position as a primary piscivore makes them suitable for Hg monitoring (Wren and MacCrimmon 1986). More importantly, our results suggest that Hg contamination of walleyes and other sport fishes in Prairie Pothole lakes should be monitored regularly, particularly after lake levels increase. Lake

surface area change may prove to be a reliable predictor of Hg concentrations, which would be useful for identifying lakes with a potential risk of Hg contamination.

Acknowledgments We extend a special thanks to R. Hanten (South Dakota Department of Game, Fish & Parks), P. Snyder and S. Mine-rich (South Dakota Department of Environment and Natural Resources) for technical assistance and providing much of the data used in the study. S. Stukel and M. Brown provided environmental data for a subset of lakes ($n=10$) used in the study. Thanks to M. Bouchard for assistance in acquiring and digitizing surface area images. Funding for this project was provided by South Dakota State University and South Dakota Department of Game, Fish, and Parks through Federal Aid in Sport Fish Restoration Project F-15-R 1502.

References

- Bodaly RA, Rudd JWM, Fudge RJP, Kelly CA (1993) Mercury concentrations in fish related to size of remote Canadian Shield lakes. *Can J Fish Aquat Sci* 50:980–987
- Bowles K, Apte S, Maher W, Bluhdorn D (2003) Mercury cycling in Lake Gordon and Lake Pedder, Tasmania (Australia) I: in-lake processes. *Water Air Soil Pollut* 147:3–23
- Collin-Hansen C, Anderson R A, Steinnes E (2005) Molecular defense systems are expressed in the king bolete (*Boletus Edulis*) growing near metal smelters. *Mycologia* 97:973–983
- Edwards SC, MacLeod CL, Lester JN (1999) Mercury contamination of the eel (*Anguilla anguilla*) and roach (*Rutilus rutilus*) in East Anglia, UK. *Environ Monit Assess* 55:371–387
- EPA (2005) EPA's Roadmap for Mercury. EPA-HQ-OPPT-2005-0013. <http://www.epa.gov/hg/pdfs/FINAL-Mercury-Roadmap-6-29.pdf>. Cited July 2006
- Essington TE, Houser JN (2003) The effect of whole-lake nutrient enrichment on mercury concentrations in age-1 yellow perch. *Trans Am Fish Soc* 132:57–68
- Gilmour C, Krabbenhoft D, Orem W, Aiken G (2004) Everglades Consolidated Report. Appx. 2b-1. <http://www.sfwmd.gov/org/ema/everglades>
- Gossman (2003) Mercury Deposition Model vs. Measurements. Gossman Consulting Inc:8(4):1–3. <http://www.gcisolutions.com/gcitrn0403.htm>
- Grieb TM, Discoll TM, Gloss CT, Schofield CP, Bowie GL, Porcella DB (1990) Factors affecting mercury accumulation in fish in the upper Michigan peninsula. *Environ Toxicol Chem* 9:919–930
- Hakanson L (2003) Consequences and correctives related to lake acidification, liming and mercury in fish - A case-study for Lake Hultesjön, Sweden, using the Lake Web-model. *Environ Mod Assess* 8:275–283
- Jackson TA (1988) The mercury problem in recently formed reservoirs of northern Manitoba (Canada): effects of impoundment and other factors on the production of methyl mercury by microorganisms in sediments. *Can J Fish and Aquat Sci* 45:97–121
- Jones RD, West-Thomas J, Arfstrom C (1997) Closed-ampule digestion procedure for the determination of mercury in soil and tissue using cold vapor atomic fluorescence spectrometry. *Bull Environ Contam Toxicol* 59:29–34
- MacCrimmon HR, Wren CD, Gots BL (1983) Mercury uptake by lake trout, *Salvelinus namaycush*, relative to age, growth, and diet in Tadenac Lake with comparative data from other Precambrian shield lakes. *Can J Fish and Aquat Sci* 40:114–120
- McMurtry MJ, Wales DL, Scheider WA, Beggs GL, Dimond PE (1989) Relationship of mercury concentrations in lake trout

- (*Salvelinus namaycush*) and smallmouth bass (*Micropterus dolomieu*) to the physical and chemical characteristics of Ontario lakes. *Can J Fish Aquat Sci* 46:426–434
- Norstrom RJ, McKinnon AE, De Freitas ASW (1976) A bioenergetics-based mode for pollutant accumulation by fish. Simulation of PCB and methylmercury residue levels in Ottawa River yellow perch (*Perca fluvialis*). *J Fish Res Board Can* 33:248–267
- Olsson M (1976) Mercury level as a function of size and age in northern pike, one and five years after the mercury ban in Sweden. *Ambio* 5:73–76
- Pickhardt PC, Folt CL, Chen CY, Klaue B, Blum JD (2002) Algal blooms reduce the uptake of toxic methylmercury in freshwater food webs. *Ecology* 99:4419–4423
- Porvari P (1998) Sources and fate of mercury in aquatic ecosystems. *Sci Total Environ* 213:279–290
- Rada RG, Wiener JG, Winfrey MR, Powell DE (1989) Recent increases in atmospheric deposition of mercury to north-central Wisconsin lakes inferred from sediment analysis. *Arch Environ Contam Toxicol* 18:175–181
- Rodgers DW, Qadri SU (1982) Growth and mercury accumulation in yearling yellow perch, *Perca flavescens*, in the Ottawa River, Ontario. *Env Biol Fish* 7:377–383
- Rosenberry DO (2003) Climate of the Cottonwood Lake Area. In: Hydrological, chemical, and biological characteristics of a prairie pothole wetland complex under highly variable climate conditions- The Cottonwood Lake Area, East-Central North Dakota. US Geological Survey Professional Paper 1675
- Rudd JWM (1995) Sources of methyl mercury to freshwater ecosystems: a review. *Water Air Soil Pollut* 80:697–713
- Snodgrass JW, Jagoe CH, Bryan AL, Brant HA, Burger J (2000) Effects of trophic status and wetland morphology, hydroperiod, and water chemistry on mercury concentrations in fish. *Can J Fish Aquat Sci* 57:171–180
- Sorensen JA, Glass GE, Schmidt KW (1994) Regional patterns of wet mercury deposition. *Environ Sci Technol* 28:2025–2032
- Sorensen JA, Kallemeyn LW, Sydor M (2005) Relationship between mercury accumulation in young-of-the-year yellow perch and water-level fluctuations. *Environ Sci Technol* 39:9237–9243
- South Dakota Game, Fish, Parks (2006) Current South Dakota Fish Consumption Advisories. <http://www.state.sd.us/doh/Fish/index.htm> cited 28 September 2006
- St. Louis VL, Rudd JWM, Kelly CA, Bodaly RA, Paterson MJ, Beaty KG, Hesslein RH, Heyes A, Majewski AR (2004) The rise and fall of mercury methylation in an experimental reservoir. *Environ Sci Technol* 38:1348–1358
- Stukel SM (2003) Assessing the sustainability of fish communities in glacial lakes: habitat inventories and relationships between lake attributes and fish communities. Unpublished M.S. Thesis, South Dakota State University, Brookings
- Suedel BC, Boraczek JA, Peddicord PK, Clifford PA, Dillon TM (1994) Trophic transfer and biomagnification potential of contaminants in aquatic ecosystems. *Rev Environ Contam Toxicol* 136:21–89
- Swain EB, Engstrom DR, Brigham ME, Henning TA, Brezonik PL (1992) Increasing rates of atmospheric mercury deposition in midcontinental North America. *Science* 257:784–787
- Verta M (1990) Changes in fish mercury concentrations in an intensively fished lake. *Can J Fish Aquat Sci* 47:1888–1897
- Wren CD, MacCrimmon HR (1983) Mercury levels in the Sunfish, *Lepomis gibbosus*, relative to pH and other environmental variables of precambrian shield lakes. *Can J Fish Aquat Sci* 40:1737–1744
- Wren CD, MacCrimmon HR (1986) Comparative bioaccumulation of mercury in two adjacent freshwater ecosystems. *Water Res* 20:763–769
- Yu LP (2005) Cloud point extraction preconcentration prior to high-performance liquid chromatography for speciation analysis of mercury in fish samples. *J Agric Food Chem* 53:9656–9662